

Enantioselective Synthesis of (+) and (-)-cis-3-Aminocyclopentanecarboxylic acids by Enzymatic Asymmetrization

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Abstract: Both enantiomers of cis-3-aminocyclopentanecarboxylic acid (GABA analogs, inhibitory neurotransmitter) have been prepared via enzymatic asymmetrization of cis-1,3-cyclopentanedicarboxylic acid.

All four stereoisomers of 3-aminocyclopentanecarboxylic acid have a potent action on mammalian central nervous system. These compounds are conformationally restricted analogues of the inhibitory neurotransmitter γ -aminobutyric acid (GABA) and they are useful stereomeric probes of the GABA binding sites topography¹. This potent activity of 3-aminocyclopentanecarboxylic acids contrasts with the much weaker effects of the corresponding 3-aminocyclohexanecarboxylic acids. We report here a chemoenzymatic synthesis of both enantiomers of cis-3-aminocyclopentanecarboxylic acid.

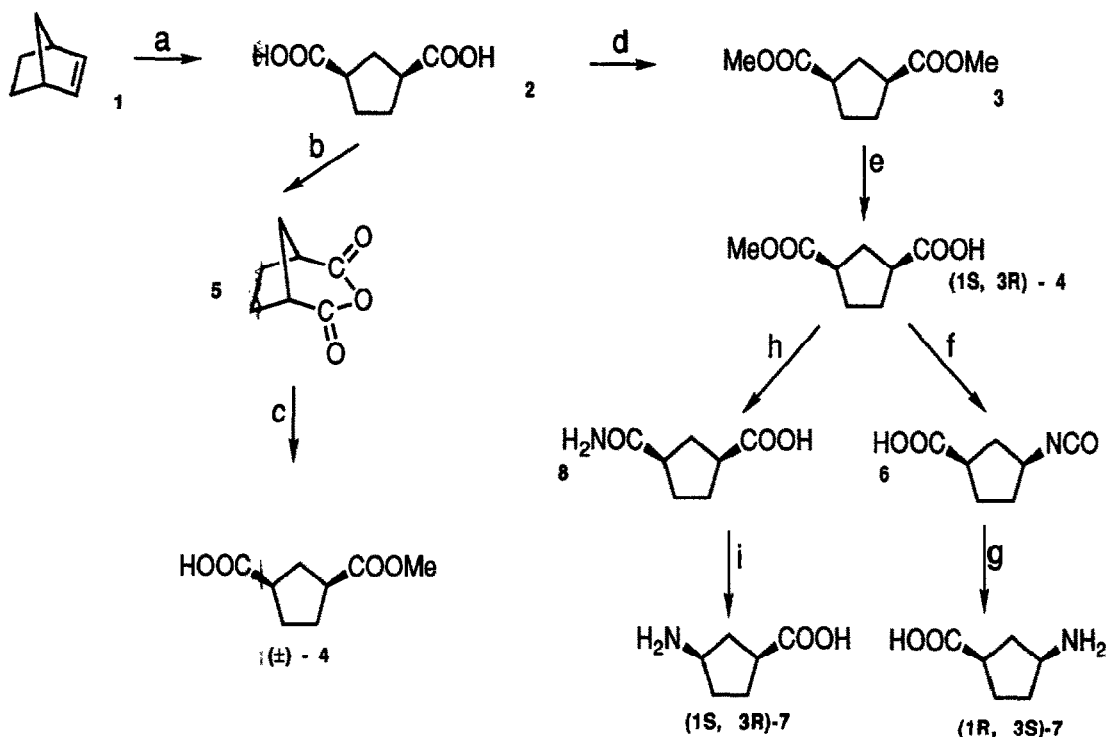
The previously reported routes to the 3-aminocyclopentanecarboxylic acids involved separation of cis/trans mixtures and resolution of enantiomers². The key feature of the present approach is an enzymic discrimination of enantiotopic ester groups of the meso cis-1,3-cyclopentanedicarboxylic acid **2**. Ozonolysis of norbornylene **1**, followed by treatment with hydrogen peroxide gave diacid **2** (scheme 1). Esterification of diacid **2** with methanol in the presence of an acidic resin as a catalyst gave diester **3**. Pig liver esterase (PLE) catalyzed hydrolysis of diester **3** has been reported³ but the enantiomeric purity of the product (mono-ester **4**) was low (*ee* = 34%). We did some preliminary screening to find suitable enzymes for the enantiotopic hydrolysis of meso-**3**. Among various proteases, lipases and esterases tested, it was found that both cholesterol esterase (CE) and subtilisin Carlsberg (SC) give the (1*S*, 3*R*) mono-ester **4** in high chemical and optical yield (SC : 85% yield, *ee* = 88%; CE : 95% yield, 90% *ee*). Racemic **4** obtained by dehydration of **2** with acetic anhydride followed by reaction of anhydride **5** with methanol was employed as a reference compound in NMR experiments.

The absolute configuration of **4** was determined by comparison of the specific rotation with reported values³. The enantiomeric purity of **4** was measured by reaction with (S)-(+)-1-(1-naphthyl) ethyl amine in the presence of 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (EDC) followed by ¹H-NMR (200 MHz) analysis of the resulting diastereoisomeric amides. The carboxyl group of (1*S*, 3*R*)-**4** was converted to an amino group with retention of configuration through a Curtius rearrangement; thus, the mono-ester was first treated with ethyl chloroformate at 0°C in acetone and with sodium azide. Acetone was replaced by toluene and the solution was heated to 100°C and the resulting isocyanate **6** was hydrolysed with aqueous HCl to give (1*R*, 3*S*)-**7**: [α]_D²⁰ -6.4 (C 1, H₂O); lit.^{2b} [α]_D²⁰ -7 (C 1, H₂O). On the other hand, ammonolysis of **4** in a pressure tube produced the corresponding amide **8** which was submitted to an Hofmann rearrangement with bis(trifluoroacetoxy)iodobenzene to give (1*S*, 3*R*)-**7**: [α]_D²⁰ +6.4 (C 1, H₂O); lit.^{2b} [α]_D²⁰ +6 (C 1, H₂O).

The enantiomeric purity of (-)-**7** and (+)-**7** was checked by NMR analysis of diastereoisomeric N-trifluoroacetyl amides obtained by reaction with (R)-1-(1-naphthyl) ethyl amine.

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Scheme 1



Reagents and Conditions: (a) *i*) O₃, MeOH, -78°C, *ii*) H₂O₂ 30%, HCOOH, 50°C, (93%); (b) Acetic anhydride, 120°C, (83%); (c) MeOH, reflux, (94%); (d) MeOH, Dowex® H⁺ resin, reflux (91%); (e) cholesterol esterase, phosphate buffer pH 7.0, 37°C, MeCN 1%, (95%); (f) *i*) Ethyl chloroformate, Et₃N, acetone, -5°C, *ii*) NaN₃, H₂O, -5°C, *iii*) toluene, reflux, (74%) (g) HCl 8N, MeOH 10%, (80%); (h) NH₃, MeOH, pressure, (82%) (i) Bis(trifluoroacetoxy)iodobenzene, MeCN, H₂O, (92%)

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